ORGANIC LETTERS

2009 Vol. 11, No. 1 205-208

An Efficient and General Enantioselective Synthesis of Sphingosine, Phythosphingosine, and 4-Substituted Derivatives[†]

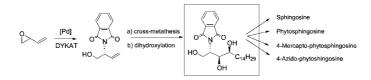
Josep Llaveria, Yolanda Díaz, M. Isabel Matheu,* and Sergio Castillón*

Departament de Química Analítica i Química Orgànica, Facultat de Química, Universitat Rovira i Virgili, C/Marcel.lí Domingo s/n, 43007 Tarragona, Spain

sergio.castillon@urv.cat; maribel.matheu@urv.cat

Received October 15, 2008

ABSTRACT



A general and efficient protocol for the enantioselective synthesis of sphingosine, phythosphingosine, and 4-substituted derivatives was established. These compounds were obtained from a common intermediate prepared from butadiene monoepoxide by a synthetic sequence involving enantioselective allylic substitution, cross-metathesis, and dihydroxylation.

Sphingolipids are important structural and functional components of the plasma membranes of essentially all eukaryotic cells. They play critical roles in many physiological processes, including immune response, cell recognition, adhesion, and apoptosis. Recent studies implicate sphingolipids in many of the most common human diseases, including diabetes, cancers, infection by microorganisms, Alzheimer's disease, heart disease, and an array of neurological syndromes. He most important sphingolipids are sphingosine and phytosphingosine, which when acylated with a fatty acid and glycosylated with galactose produce galac-

tosylceramide (GalCer) and α -GalCer (KRN7000), respectively (Figure 1). Recently, structurally modified sphin-

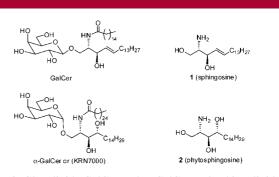


Figure 1. Glycolipids GalCer and α -GalCer and sphingolipids.

† Dedicated to Professor Josep Font on occasion of his 70th birthday. (1) (a) Riethmüller, J.; Riehle, A.; Grassmé, H.; Gulbins, E. *Biochim. Biophys. Acta* **2006**, *1758*, 2139–2147. (b) Snook, C. F.; Jones, J. A.; Hannun, Y. A. *Biochim. Biophys. Acta* **2006**, *1761*, 927–946.

(2) Summers, S. A.; Nelson, D. H. Diabetes 2005, 54, 591-602.

gosines⁷ and phytosphingosines^{8,9} have attracted more attention because some of their analogues have been observed to introduce morphological changes in neuronal cells¹⁰ and behave as enzyme inhibitors.¹¹

⁽³⁾ Modrak, D. E.; Gold, D. V.; Goldenberg, D. M. *Mol. Cancer Ther.* **2006**, *5*, 200–208.

⁽⁴⁾ Heung, L. J.; Luberto, C.; Del Poeta, M. Infect. Immun. 2006, 74, 28–39.

⁽⁵⁾ Zhou, S.; Zhou, H.; Walian, P. J.; Jap, B. K. Biochemistry 2007, 46, 2553–2563.

⁽⁶⁾ Kolter, T.; Sandhoff, K. Biochim. Biophys. Acta 2006, 1758, 2057–2079.

Since sphingosine and its derivatives are available only in limited amounts from natural sources and because of purity requirements for biological testing, there is a growing interest in developing efficient methods for their synthesis. ¹² These compounds have been synthesized by various routes, but primarily from compounds of the chiral pool, particularly amino acids (L-serine)¹³ and carbohydrates. ¹⁴ Asymmetric syntheses based on the use of chiral auxiliaries, such as sulfoxides, ¹⁵ chiral aziridines, ¹⁶ or chiral sulfur ¹⁷ and nitrogen ¹⁸ ylides, or on catalytic procedures, such as Sharpless asymmetric epoxidation ¹⁹ and dihydroxylation reactions, ^{16,20} the aldol reaction, ²¹ and organocatalytic procedures, have also been described. ²²

Recently, we reported efficient procedures for the glycosylation of ceramides that facilitated the synthesis of GalCer²³ and KRN 7000.²⁴ New analogues of these compounds containing structural modifications in the sphingolipid moiety have been reported very recently.²⁵ In this work, we describe

(7) Sawatzki, P.; Kolter, T. Eur. J. Org. Chem. 2004, 3693–3700, and references therein.

(8) The four stereoisomers of phytosphingosine were prepared from the natural product. Kim, S.; Lee, N.; Lee, S.; Lee, T.; Lee, Y. M. J. Org. Chem. 2008, 73, 1379–1385.

(9) Oldendorf, J.; Haufe, G. Eur. J. Org. Chem. 2006, 4463-4472.

(10) Van Echten-Deckert; Zschosche, A.; Bär, T.; Schmidt, R. R.; Raths,
A.; Heinemann, T.; Sandhoff, K. J. Biol. Chem. 1997, 272, 15825–15833.
(11) Brodesser, S.; Sawatzki, P.; Kolter, T. Eur. J. Org. Chem. 2003, 2021–2024.

(12) (a) Koskinen, P. M.; Koskinen, A. M. P. *Synthesis* **1998**, 1075–1091. (b) Howell, A. R.; Ndakala, A. J. *Curr. Org. Chem.* **2002**, *6*, 365–391.

(13) (a) Chun, J.; Lee, G.; Byun, H.-P.; Bittman, R. *Tetrahedron Lett.* **2002**, *43*, 375–377. (b) Yadav, J. S.; Geetha, V.; Raju, A. K.; Gnaneshwar, D.; Chandrasekhar, S. *Tetrahedron Lett.* **2003**, *44*, 2983–2985. (c) Lombardo, M.; Capdevila, M. G.; Pasi, F.; Trombini, C. *Org. Lett.* **2006**, *8*, 3303–3305. (d) Lee, J.-M.; Lim, H.-S.; Chung, S.-K. *Tetrahedron: Asymmetry* **2002**, *13*, 343–347. (e) Yamamoto, T.; Hasegawa, H.; Hakogi, T.; Katsumura, S. *Org. Lett.* **2006**, *8*, 5569–5572. (f) Masuda, Y.; Mori, K. *Eur. J. Org. Chem.* **2005**, 4789–4800. (g) Duffin, G. R.; Ellames, G. J.; Hartmann, S.; Herbert, J. M.; Smith, D. I. *J. Chem. Soc., Perkin Trans. 1* **2000**, 2237–2242. (h) Mori, K.; Masuda, Y. *Tetrahedron Lett.* **2003**, *44*, 9197–9200.

(14) (a) Luo, S.-Y.; Thopate, S. R.; Hsu, C.-Y.; Hung, S.-C. Tetrahedron Lett. 2002, 23, 4889-4892. (b) Chaudhari, V. D.; Kumar, K. S. A.; Dhavale, D. D. Org. Lett. 2005, 7, 5805–5807. (c) Lin, C.-C.; Fan, G.-T.; Fan, J.-M. Tetrahedron Lett. 2003, 44, 5281-5283. (d) Chiu, H.-Y.; Tzou, D.-L. M.; Patkar, L. N.; Lin, C.-C. *J. Org. Chem.* **2003**, *68*, 5788–5791. (e) Plettenburg, O.; Bodmer-Narkevich, V.; Wong, C.-H. *J. Org. Chem.* **2002**, 67, 4559–4564. (f) Graziani, A.; Passancatelli, P.; Piancatelli, G.; Tani, S. Tetrahedron: Asymmetry 2000, 11, 3921-3937. (g) Wild, R.; Schmidt, R. R. Tetrahedron: Asymmetry 1994, 5, 2195-2208. (h) Fan, G.-T.; Pan, Y.-S.; Lu, K.-C.; Cheng, Y.-P.; Lin, W.-C.; Lin, S.; Lin, C.-H.; Wong, C.-H.; Fang, J.-M.; Lin, C.-C. Tetrahedron 2005, 61, 1855-1862. (i) Figueroa-Pérez, S.; Schmidt, R. R. Carbohydr. Res. 2000, 328, 95-102. (j) Compostella, F.; Franchini, L.; De Libero, G.; Palmisano, G.; Ronchetti, F.; Panza, L. Tetrahedron 2002, 58, 8703–8708. (k) Duclos, R. I., Jr. Chem. Phys. Lipids 2001, 111, 111-138. (l) Milne, J. E.; Jarowickki, K.; Kocienski, P. J.; Alonso, J. Chem. Commun. 2002, 426-427. (m) Van den Berg, R. J. B. H. N.; Korevaar, C. G. N.; van der Marel, G. A.; Overkleeft, H. S.; van Boom, J. H. Tetrahedron Lett. 2002, 43, 8409-8412.

(15) Zhong, Y.-W.; Dong, Y.-Z.; Fang, K.; Izumi, K.; Xu, M.-H.; Lin, G.-Q. J. Am. Chem. Soc. 2005, 127, 11956–11957.

(16) Yoon, H. J.; Kim, Y.-W.; Lee, B. K.; Lee, W. K.; Kim, Y.; Ha, Y.-J. Chem. Commun. **2007**, 79–81.

(17) Morales-Serna, J. A.; Llaveria, J.; Díaz, Y.; Matheu, M. I.; Castillón, S. Org. Biomol. Chem. 2008, doi 10.1039/b814882a.

(18) Disadee, W.; Ishikawa, T. J. Org. Chem. 2005, 70, 9399-9406.

(19) (a) Olofsson, B.; Somfai, P. *J. Org. Chem.* **2003**, *68*, 2514–2517. (b) Torssell, S.; Somfai, P. *Org. Biomol. Chem.* **2004**, *2*, 1643–1646. (c) Righi, G.; Ciambrone, S.; D'Achille, C.; Leonelli, A.; Bonini, C. *Tetrahedron* **2006**, *62*, 11821–11826.

(20) (a) Martin, C.; Prünk, W.; Bortolussi, M.; Bloch, R. *Tetrahedron: Asymmetry* **2000**, *11*, 1585–1592. (b) He, L.; Byun, H.-S.; Bittman, R. *J. Org. Chem.* **2000**, *65*, 7618–7626.

a new and efficient enantioselective method for synthesizing sphingosine (1), phytosphingosine (2), and new 4-substituted derivatives (3, 4) (Scheme 1) partially protected. In the

Scheme 1. Retrosynthesis

proposed retrosynthesis, compounds 1–4 can be obtained from a common intermediate 5 (Scheme 1). Nucleophilic substitution at position 4 in 5 must allow the introduction of different substituents, affording the natural product and derivatives. Compound 5 can be obtained by the dihydroxylation of compound 6, which in turn can be synthesized from compound 7 by a cross-metathesis reaction. The main advantage of this strategy is its high versatility, allowing the synthesis of not only sphingosine and phytosphingosine but also a range of structural analogues from a common precursor.

Chiral synthon 7 (NR₂ = phthalimido) was obtained by a palladium-catalyzed dynamic kinetic asymmetric transformation (DYKAT) from the racemic butadiene monoepoxide (8)²⁶ (Scheme 2).

Scheme 2. Synthesis of Alkene 6

Initially we explored the cross metathesis reaction using the second generation Grubbs catalyst, which is compatible with a wide range of functionalities. 13e,14b,17,19b In preliminary screening experiments, compound **7** was reacted with a 2-fold excess of 1-hexadecene (**9**) in refluxing dichloromethane to afford **6** at a 82% yield and an E/Z ratio of 18:1 (Scheme 2). Since the metathesis reaction proceeds under thermodynamic control, both the yield and stereose-

206 Org. Lett., Vol. 11, No. 1, 2009

lectivity can be improved by increasing the 9/7 ratio and the reaction time. In this way, using 4 equiv of 9 and maintaining the reaction for 12 h, compound 6 was obtained in an quantitative yield, and the E isomer was exclusively detected by NMR.

Compound 6 was then reacted with OsO₄/NMO to obtain a mixture of compounds 5 and 10 in an almost quantitative yield in a ratio of 3.3:1 (Scheme 3) (entry 1, Table 1).

Scheme 3. Dihydroxylation of Alkene 6

Table 1. Dihydroxylation of Alkene 6

entry	reagent	temp (°C)	yield (%)	ratio 5:10
1	OsO ₄ /NMO	rt	99	3.3:1
2	OsO ₄ /NMO	0	57	3.4:1
3^a	OsO_4/NMO	-78	95	3.4:1
4^b	OsO ₄ /NMO	-78	93	3.8:1
5	AD-MIX α^c	rt		
6	AD-MIX β^d	rt		
7	$[\mathrm{Os}]/(\mathrm{DHQ})_2\mathrm{PYR}^e$	rt	99	5.1:1

 a OsO₄ (1 equiv) and TMDA (1.1 equiv) were used. b TEEDA was used as the ligand. c Ligand (DHQ)₂PHAL. d Ligand (DHQD)₂PHAL. e K₂OsO₂(OH)₄ (0.02 equiv), (DHQ)₂PYR (0.03 equiv), CH₃SO₂NH₂ (1.2 equiv), K₂CO₃ (0.03 equiv), NaHCO₃ (0.03 equiv), K₃Fe(CN)₆, (0.03 equiv).

Decreasing the temperature had a negative effect on the yield and no effect on the stereoselectivity (entry 2). An attempt was made to increase the stereoselectivity by carrying out the reaction at -78 °C and using stoichiometric amounts of OsO₄ in the presence of different diamine ligands. When

tetramethylethylenediamine (TMEDA) was used, the steroselectivity was similar to that reported in entry 1 (entry 3). The use of tetraethylethylenediamine (TEEDA) slightly increased the **5/10** ratio to 3.8:1, in an 93% yield (entry 4).

It has been reported that the asymmetric dihydroxylation reactions of related substrates afforded excellent yields and stereoselectivities of the L-*lyxo* and D-*xylo* phytosphingosines, using AD-MIX α and β, respectively;^{27,28} however, when compound 6 was treated with commercial AD-MIX mixtures,²⁷ no reaction was observed (entries 5, 6, Table 1). The reaction was attempted using a freshly prepared mixture of [K₂OsO₂(OH)₄] and [K₃Fe(CN)₆] in the presence of ligands (DHQD)₂-PHAL or (DHQ)₂-PHAL, in 'BuOH/H₂O (1:1), but unfortunately, the starting material was again exclusively recovered. Finally, in the presence of K₂OsO₂(OH)₄]/[K₃Fe(CN)₆]/(DHQ)₂PYR, compounds 5/10 were obtained in a quantitative yield with a ratio of 5.1:1 (Entry 7).

With compound **5** in hand, the next step involved the selective protection of the hydroxyl groups at positions 1 and 3 and the activation of the 4-OH as a leaving group. We initially attempted the simultaneous protection of 1- and 3-OH by reaction with 'Bu₂Si(OTf)₂ and further activation of the 4-OH as a triflate. However, the subsequent elimination provided a very poor yield of the sphingosine derivative. Alternatively, **5** was reacted with TBDPSCl, affording compound **11** in an 89% yield, which was then treated with thionyl chloride and RuO₄/NaIO₄, affording sulfate **12** in a quantitative yield (Scheme 4).

Scheme 4. Synthesis of Key Intermediate 12 and Sphingosine

Compound **12** was then reacted with DBU in the presence of tetrabutylammonium iodide to obtain compound **13** in an 82% yield. Further deprotection of **13** by reaction with TBAF in THF at room temperature and treatment with hydrazine afforded sphingosine (**1**) in an 82% yield.

Similarly, **12** was also reacted with benzoic acid and Cs₂CO₃, to produce compound **14** in a 91% yield (Scheme 5). This excellent regioselectivity was also observed for other nucleophiles and was atributed to the steric and electronic interactions between neighboring sustituents and nucleophiles. ⁸ Compound **14** was also deprotected by reacting it

Org. Lett., Vol. 11, No. 1, 2009

^{(21) (}a) Cai, Y.; Ling, C.-C.; Bundle, D. R. *Org. Biomol. Chem.* **2006**, *4*, 1140–1146. (b) Kobayashi, J.; Nakamura, M.; Mori, Y.; Yamashita, Y.; Kobayashi, S. *J. Am. Chem. Soc.* **2004**, *126*, 9192–9193.

⁽²²⁾ Enders, D.; Palecek, J.; Grondal, C. Chem. Commun. 2006, 655–657.

^{(23) (}a) Morales-Serna, J. A.; Boutureira, O.; Díaz, Y.; Matheu, M. I.; Castillón, S. *Org. Biomol. Chem.* **2008**, *6*, 443–445. (b) Morales-Serna, J. A.; Díaz, Y.; Matheu, M. I.; Castillón, S. *Org. Biomol. Chem.* **2008**, *6*, 3831–3836.

⁽²⁴⁾ Boutureira, O.; Morales-Serna, J. A.; Díaz, Y.; Matheu, M. I.; Castillón, S. *Eur. J. Org. Chem.* **2008**, 1851–1854.

^{(25) (}a) Wipf, P.; Pierce, J. G. *Org. Lett.* **2006**, *8*, 3375–3378. (b) Dere, R. T.; Zhu, X. *Org. Lett.* **2008**, *10*, 4641–4644. (c) Leung, L.; Tomassi, C.; van Beneden, K.; Decry, T.; Elewaut, D.; Elliott, T.; Al-Shamkhani, A.; Ottensmeier, C.; Van Calenberg, S.; Werner, J.; Williams, T.; Linclau, B. *Org. Lett.* **2008**, *10*, 4433–4436.

^{(26) (}a) Trost, B. M.; Bunt, R. C.; Lemoine, R. C.; Calkins, T. L. J. Am. Chem. Soc. **2000**, 122, 5968–5976. (b) Trost, B. M.; Horne, D. B.; Woltering, M. J. Angew. Chem., Int. Ed. **2003**, 42, 5987–5990.

Scheme 5. Synthesis of Phytosphingosine (2)

with TBAF and hydrazine to furnish phytosphingosine (2) in an 89% yield. NMR spectra and optical rotation of compounds $\mathbf{1}^{19b}$ and $\mathbf{2}^{29}$ match the reported values for the natural products.

The possibility to obtain analogues of phytosphingosine modified at position 4 was illustrated by synthesizing the new 4-mercapto and the 4-azido derivatives (Scheme 6).

Scheme 6. Synthesis of Phytosphingosine Derivatives 15 and 16

Thus, compound 12 was reacted with BzSH and Cs_2CO_3 to render compound 15 in an 87% yield. In a parallel experiment, compound 12 was reacted with sodium azide in the presence of catalytic 15-crown-5 to afford compound 16 in an 89% yield.

In conclusion, D-*erythro*-sphingosine (1), *N*-phtalimido-D-*lyxo*- (5), D-*ribo*-phytosphingosine (2), and 4-mercapto (15) and 4-azido (16) analogs were prepared by a highly efficient and enantioselective procedure (Scheme 7). This procedure

Scheme 7. Synthesis of Compounds 1, 2, 5, 15 and 16 from 7

starts from butadiene monoepoxide and uses a Pd-catalyzed DYKAT process, a cross-metathesis using a second generation Grubbs catalysis and a dihydroxylation reaction to produce the key intermediate 5. From this intermediate, the target compounds were obtained by protection, substitution, or elimination of 4-OH and deprotection. This procedure is the most efficient for preparing 1 and 2 using asymmetric synthesis procedures and opens the way for preparing a large variety of 4-phytosphingosine derivatives.

Acknowledgment. The authors acknowledge the financial support of DGESIC, CTQ2005-03124 and Consolider Ingenio 2010, CSD2006-0003 (Ministerio de Educación, Spain). We thank Servei de Recursos Científics (URV) for technical assistance. Fellowships from DURSI (Generalitat de Catalunya) and Fons Social Europeu to J.L. are also acknowledged. We also thank Dr. Christopher Cobley (Dow Pharma) for the generous gift of a sample of the starting material.

Supporting Information Available: General experimental methods, experimental procedures, compound characterization data, and NMR spectra for all new compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

OL802379B

Org. Lett., Vol. 11, No. 1, 2009

⁽²⁷⁾ Kolb, H. C.; VanNieuwenhze, M. S.; Sharpless, K. B. Chem. Rev. **1994**, 94, 2483–2547.

^{(28) (}a) Imashiro, R.; Sakurai, O.; Yamashita, T.; Horikawa, H. *Tetrahedron* **1998**, *54*, 10657–10670. (b) Mormeneo, D.; Casas, J.; Llebaria, A.; Delgado, A. *Org. Biomol. Chem.* **2007**, *5*, 3769–3777.

⁽²⁹⁾ Dondoni, A.; Fantin, G.; Fogagnolo, M.; Pedrini, P. J. Org. Chem. 1990, 55, 1439–1446.